Monatshefte für Chemie Chemical Monthly Printed in Austria

# Experimental Evidence for a Valinomycin – Proton Complex

Emanuel Makrlík<sup>1</sup> and Petr Vaňura<sup>2,\*</sup>

<sup>1</sup> Faculty of Applied Sciences, University of West Bohemia, 306 14 Pilsen, Czech Republic
 <sup>2</sup> Prague Institute of Chemical Technology, 166 28 Prague 6, Czech Republic

Received June 17, 2005; accepted (revised) July 5, 2005 Published online January 20, 2006 © Springer-Verlag 2006

Summary. From extraction experiments and  $\gamma$ -activity measurements, the extraction constant corresponding to the equilibrium  $H^+(aq) + NaL^+(nb) = HL^+(nb) + Na^+(aq)$  taking place in the twophase water-nitrobenzene system (*L*=valinomycin, aq = aqueous phase, nb = nitrobenzene phase) was evaluated as log  $K_{ex}(H^+, NaL^+) = -1.1 \pm 0.1$ . Further, the stability constant of the valinomycinproton complex in nitrobenzene saturated with water was calculated as log  $\beta_{nb}(HL^+) = 5.3 \pm 0.1$ . Finally, the stability constants of complexes of some univalent cations with valinomycin were summarized and discussed.

Keywords. Antibiotics; Macrocycles; Protonation; Stability constant; Valinomycin.

## Introduction

The antibiotic valinomycin was discovered in cultures *Streptomyces fulvissimus* [1]. It is a macrocyclic depsipeptide exhibiting threefold symmetry (see Fig. 1) [2]. It forms complexes with alkali metal ions. The stabilities of these complexes show a pronounced dependence on the ion radius [2–5]. Because of the lipophilic outer envelope of the complex, valinomycin enables univalent cations to be transported across membranes of cells and cell organelles [3], and consequently, it is a powerful decoupler of oxidative phosphorylation in mitochondria [6]. It also gives rise to ion-selective membrane potentials at bilayer lipid membranes [3]. The very large difference in the stabilities of the potassium and the sodium complexes is the cause of the high potassium selectivity of the valinomycin-based ion-selective electrode [7].

Up to now, a complex species of the proton with valinomycin has not been proved. In the present communication, the stability of protonated valinomycin in nitrobenzene saturated with water was evaluated.

<sup>\*</sup> Corresponding author. E-mail: Petr.Vanura@vscht.cz

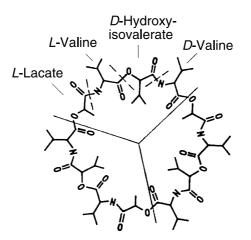


Fig. 1. Formula of valinomycin

#### **Results and Discussion**

With regard to the previous results [9, 10] the two-phase water-HCl/nitrobenzene-NaDCC extraction system can be described by the following equilibrium (1) with the corresponding exchange extraction constant  $K_{ex}(H^+, Na^+)$ ; aq and nb denote the presence of the species in the aqueous and nitrobenzene phases.

$$H^{+}(aq) + Na^{+}(nb) = H^{+}(nb) + Na^{+}(aq); \qquad K_{ex}(H^{+}, Na^{+})$$
(1)

For the constant  $K_{ex}(H^+, Na^+)$  it can be written [9] as shown by Eq. (2) where  $K_{H^+}^i$  and  $K_{Na^+}^i$  are the individual extraction constants for H<sup>+</sup> and Na<sup>+</sup> in the waternitrobenzene system [9].

$$\log K_{\rm ex}({\rm H}^+,{\rm Na}^+) = \log K_{{\rm H}^+}^i - \log K_{{\rm Na}^+}^i \tag{2}$$

Knowing log  $K_{\text{H}^+}^i = -5.7$  [9] and log  $K_{\text{Na}^+}^i = -6.0$  [9], the exchange extraction constant  $K_{\text{ex}}(\text{H}^+, \text{Na}^+)$  was simply calculated from Eq. (2) as log  $K_{\text{ex}}(\text{H}^+, \text{Na}^+) = 0.3$ .

In terms of previous papers [9–12], the two-phase water-HCl/nitrobenzene-NaDCC-L (L = valinomycin) extraction system, chosen for determination of stability of the complex HL<sup>+</sup>, can be characterized by the main chemical equilibrium (3) to which the equilibrium extraction constant as shown by Eq. (4) corresponds.

$$\mathbf{H}^{+}(\mathbf{aq}) + \mathbf{Na}L^{+}(\mathbf{nb}) = \mathbf{H}L^{+}(\mathbf{nb}) + \mathbf{Na}^{+}(\mathbf{aq}); \qquad K_{\mathrm{ex}}(\mathbf{H}^{+}, \mathbf{Na}L^{+})$$
(3)

$$K_{\rm ex}({\rm H}^+,{\rm Na}L^+) = \frac{[{\rm H}L^+]_{\rm nb}[{\rm Na}^+]_{\rm aq}}{[{\rm H}^+]_{\rm aq}[{\rm Na}L^+]_{\rm nb}}$$
(4)

It is necessary to emphasize that valinomycin is a considerably hydrophobic ligand, practically present in the nitrobenzene phase only, where this ligand forms – with  $H^+$  and  $Na^+$  – the relatively stable complexes  $HL^+$  and  $NaL^+$ .

Following the conditions of electroneutrality in the organic and aqueous phases of Eqs. (5) and (6), the mass balances of hydrogen and sodium at equal volumes of the nitrobenzene and aqueous phases of Eqs. (7) and (8), and the measured equili-

brium distribution ratio of sodium (Eq. (9)), combined with Eq. (4) the final expression for the extraction constant (Eq. (10)) is obtained.

$$\left[\mathrm{H}L^{+}\right]_{\mathrm{nb}} + \left[\mathrm{Na}L^{+}\right]_{\mathrm{nb}} = C_{\mathrm{Na}DCC}^{\mathrm{in,nb}} \tag{5}$$

$$[H^{+}]_{aq} + [Na^{+}]_{aq} = C_{\rm HCl}^{in,aq}$$
(6)

$$\left[\mathrm{H}^{+}\right]_{\mathrm{aq}} + \left[\mathrm{H}L^{+}\right]_{\mathrm{nb}} = C_{\mathrm{HCl}}^{\mathrm{in,aq}} \tag{7}$$

$$[\mathrm{Na}^+]_{\mathrm{aq}} + [\mathrm{Na}L^+]_{\mathrm{nb}} = C_{\mathrm{Na}DCC}^{\mathrm{in,nb}}$$
(8)

$$D_{\mathrm{Na}} = [\mathrm{Na}L^+]_{\mathrm{nb}} / [\mathrm{Na}^+]_{\mathrm{aq}}$$
<sup>(9)</sup>

$$K_{\rm ex}({\rm H}^+,{\rm Na}L^+) = \frac{1}{D_{\rm Na}} \frac{C_{\rm Na}^{\rm in,nb}}{(1+D_{\rm Na})C_{\rm HCl}^{\rm in,aq} - C_{\rm Na}^{\rm in,nb}}$$
(10)

From the extraction experiments and  $\gamma$ -activity measurements by using Eq. (10), the following value of the constant  $\log K_{\text{ex}}(\text{H}^+, \text{Na}L^+)$  was evaluated:  $\log K_{\text{ex}}(\text{H}^+, \text{Na}L^+) = -1.1 \pm 0.1$ .

Moreover, with respect to Refs. [12] and [13], for the extraction constants  $K_{ex}(H^+, Na^+)$  and  $K_{ex}(H^+, NaL^+)$  defined above as well as for the stability constants of the complexes NaL<sup>+</sup> and HL<sup>+</sup> in nitrobenzene saturated with water, denoted by  $\beta_{nb}(NaL^+)$  and  $\beta_{nb}(HL^+)$ , one gets Eq. (11).

$$\log \beta_{\rm nb}(\mathrm{H}L^+) = \log \beta_{\rm nb}(\mathrm{Na}L^+) + \log K_{\rm ex}(\mathrm{H}^+, \mathrm{Na}L^+) - \log K_{\rm ex}(\mathrm{H}^+, \mathrm{Na}^+) \quad (11)$$

Using the constants  $\log K_{\text{ex}}(\text{H}^+, \text{Na}^+)$  and  $\log K_{\text{ex}}(\text{H}^+, \text{Na}L^+)$  given above, the value  $\log \beta_{\text{nb}}(\text{Na}L^+) = 6.7$  [14], and applying Eq. (11), we obtain the stability constant of the valinomycin-proton complex in nitrobenzene saturated with water at 25°C as  $\log \beta_{\text{nb}}(\text{H}L^+) = 5.3 \pm 0.1$ .

The stability constants of complexes of some univalent cations with valinomycin in methanol, acetonitrile, and nitrobenzene saturated with water are listed with the corresponding references in Table 1. From here it follows that in the media under consideration, valinomycin forms the most stable complex with  $Rb^+$  and the weakest one with Na<sup>+</sup> or H<sup>+</sup> (in nitrobenzene saturated with water) – the stability of the potassium complex is only slightly lower than that of the rubidium complex. Furthermore, the stability constants of the same metal complexes increase in the series methanol < acetonitrile < nitrobenzene saturated with water. Since the forma-

**Table 1.** Logarithms of stability constants of some univalent cations with valinomycin in methanol (a), acetonitrile (b), and nitrobenzene saturated with water (c)

Medium	$\mathrm{H}^+$	Na <sup>+</sup>	K <sup>+</sup>	$Rb^+$	Cs <sup>+</sup>
a	_	$0.7^{\mathrm{a}}$	4.5 <sup>a</sup>	$4.8^{\mathrm{a}}$	3.9 <sup>a</sup>
b	_	$2.6^{b}$	6.7 <sup>b</sup>	$6.9^{\mathrm{b}}$	$6.0^{\mathrm{b}}$
c	5.3°	6.7 <sup>d</sup>	10.4 <sup>e</sup>	$11.7^{\mathrm{f}}$	10.1 <sup>g</sup>

<sup>a</sup> Ref. [4]; <sup>b</sup> Ref. [15]; <sup>c</sup> this work; <sup>d</sup> Ref. [14]; <sup>e</sup> Ref. [16]; <sup>f</sup> Ref. [17]; <sup>g</sup> Ref. [13]

tion of an arbitrary complex is assumed to be accompanied by competition for the respective cation between solvent molecules and ligand bonding sites, the present results are obviously a consequence of the decrease in basicity and solvating power of the medium in the order methanol>acetonitrile>nitrobenzene saturated with water.

## **Experimental**

Valinomycin was supplied by Merck, Darmstadt, Germany. Cs dicarbollylcobaltate (Cs*DCC*) was synthesized in the Institute of Inorganic Chemistry, Řež, Czech Republic, using the method published by *Hawthorne et al.* [8]. The other chemicals used (Lachema, Brno, Czech Republic) were of reagent grade purity. The radionuclide <sup>22</sup>Na (DuPont, Belgium) was of standard radiochemical purity.

In order to obtain the solution of HDCC in nitrobenzene, the solution of CsDCC in this medium (0.2M) was equilibrated twice with equal volumes of 15% (v/v) n-propanol in diluted H<sub>2</sub>SO<sub>4</sub> (about 1 M). H<sub>2</sub>SO<sub>4</sub> and n-propanol were removed from the organic phase by a ten-fold equilibration with an equal volume of a solution of H<sub>2</sub>SO<sub>4</sub> in distilled H<sub>2</sub>O (about 1 M) followed by two equilibrations with distilled H<sub>2</sub>O. The concentration of HDCC in nitrobenzene was determined by neutralization titration (NaOH, bromocresol green) after a ten-fold dilution with ethanol or acetone. The equilibration of the nitrobenzene solution of HDCC with stoichiometric NaOH, which was dissolved in an aqueous solution of NaCl (0.2 M), yields the corresponding NaDCC solution in nitrobenzene.

The extraction experiments were carried out in  $10 \text{ cm}^3$  glass test-tubes covered with polyethylene stoppers:  $2 \text{ cm}^3$  of the aqueous HCl  $(1 \times 10^{-4} - 1 \times 10^{-3} M)$  and microamounts of <sup>22</sup>Na were added to  $2 \text{ cm}^3$  of the nitrobenzene solution of valinomycin and NaDCC, whose initial concentrations varied also from  $1 \times 10^{-4}$  to  $1 \times 10^{-3} M$  (in all experiments, the initial concentration of valinomycin in nitrobenzene,  $C_L^{\text{in,nb}}$ , was always equal to the initial concentration of NaDCC in this medium,  $C_{\text{NaDCC}}^{\text{in,nb}}$ . The test-tubes filled with the solutions were shaken for 2 h at  $25 \pm 1^{\circ}$ C using a laboratory shaker. Under these conditions, an equilibrium in the system under study was established after approximately 15 min. Then the phases were separated by centrifugation (2 min, 2500 rpm). Afterwards, 1 cm<sup>3</sup> samples were taken from each phase and their  $\gamma$ -activities were measured using a well-type NaI (TI) scintillation detector connected to a single channel  $\gamma$ -analyzer NK 350 (Gamma, Budapest, Hungary).

The equilibrium distribution ratio of sodium,  $D_{\text{Na}}$ , was determined as the ratio of the measured radioactivities of <sup>22</sup>Na in the nitrobenzene and aqueous samples.

### Acknowledgements

The present work was supported by the Czech Ministry of Education, Youth and Sports, Projects Nos. MSM 4977751303 and MSM 6046137307.

#### References

- [1] Brockmann H, Schmidt-Kastner G (1955) Chem Ber 88: 57
- [2] Shemyakin MM, Vinogradova EI, Feigina MYu, Aldanova NA, Loginova NF, Raybova ID, Pavlenko IA (1965) Experientia 21: 548
- [3] Shemyakin MM, Ovchinnikov YuA, Ivanov VT, Antonov VK, Vinogradova EI, Shkrob AM, Malenkov GG, Yevstratov AV, Laine IA, Melnik YuI, Ryabava ID (1969) J Membr Biol 1: 402
- [4] Funck T, Eggers F, Grell E (1972) Chimia 26: 637
- [5] Březina M, Hofmanová A, Koryta J (1974) Biophys Chem 2: 264
- [6] Shemyakin MM, Ovchinnikov YuA, Ivanov VT, Kiryuskin AA, Zhdanov GL, Ryabova ID (1963) Experientia 19: 566

- [7] Pioda LAR, Simon W, Bosshard HR, Curtius HCH (1970) Clin Chim Acta 29: 289
- [8] Hawthorne MF, Young DC, Andrews TD, Hove DV, Pilling RL, Pitts AD, Reintjes M, Warren LF, Wegner PA (1968) J Am Chem Soc 90: 879
- [9] Rais J (1971) Collect Czech Chem Commun 36: 3253
- [10] Rais J, Selucký P, Kyrš M (1976) J Inorg Nucl Chem 38: 1376
- [11] Danesi PR, Meider-Goričon H, Chiarizia R, Scibona G (1975) J Inorg Nucl Chem 37: 1479
- [12] Makrlík E, Hálová J, Kyrš M (1984) Collect Czech Chem Commun 49: 39
- [13] Makrlík E, Vaňura P (1996) J Radioanal Nucl Chem 214: 339
- [14] Makrlík E, Vaňura P (1998) ACH Models in Chemistry 135: 213
- [15] Hofmanová A, Koryta J, Březina M, Ryan TH, Angelis K (1979) Inorg Chim Acta 37: 135
- [16] Koryta J, Kozlov YuN, Skalický M (1987) J Electroanal Chem 234: 355
- [17] Daňková M, Makrlík E, Vaňura P (1997) J Radioanal Nucl Chem 221: 251